

RESEARCH ARTICLE

Azzeddine Zeraib^{1,2}
Messaoud Ramdani²
Lamia Boudjedjou²
Pierre Chalard³
Gilles Figuredo⁴

Chemical composition and antibacterial activity of *Juniperus thurifera* L. essential oils

Authors' addresses:

¹ Department of Molecular Biology,
Abbes Laghrour University,
40000 Khenchela, Algeria.

² Laboratory of Natural Resource
Valorization, Ferhat Abbas University,
19000 Setif, Algeria.

³ Blaise Pascal University, BP 10448,
F-63000 Clermont Ferrand, France.

⁴ LEXVA Analytique,
63110 Beaumont, France.

Correspondence:

Azzeddine Zeraib
Department of Biology Molecular,
Abbes Laghrour University, 40000
Khenchela, Algeria.
Tel.: +213 778 168 697
e-mail: azzeraib@yahoo.fr

Article info:

Received: 5 April 2014
Accepted: 9 June 2014

ABSTRACT

The qualitative and quantitative composition of the essential oils obtained from male and female leaves of *Juniperus thurifera* L., (growing in Algeria) has been investigated for the first time. The essential oils were obtained by hydrodistillation (0.45% from female trees and 0.53% from male trees, v/w dried material) and analyzed by gas chromatography (GC) and gas chromatography-mass spectrometry (GC-MS). Seventy-seven compounds were identified, representing more than 97% of the oils. The major components were Sabinene, α -pinene and terpinene-4-ol. The concentrations of the oil constituents: α -thujene, α -pinene, α -phellandrene, p-cymene, linalyl acetate, Δ -amorphene, germacrene D-4-ol, and 4-epi-abietal were greater in the oil of the female tree than in the oil of the male tree. Conversely, the concentrations of α -terpinene, γ -terpinene, terpinene-4-ol, elemol, α -epi-cadinol and α -eudesmol were greater in the oil of the male tree than in the oil of the female tree. However, the concentration gradient trends for both female and male trees were similar for sabinene, myrcene, linalool, β -pinene, limonene, *cis*-sabinene hydrate terpinolene, α -terpineol. The antimicrobial activity of male and female *J. thurifera* essential oils was evaluated against 14 bacteria. The results showed a variable degree of antibacterial activity depending from the type of the oil (extracted from male or female trees). Essential oils of female trees were most effective.

Key words: Cupressaceae, *Juniperus thurifera* L., sex of the trees, essential oil, antimicrobial activity

Introduction

The genus *Juniperus* L. (Cupressaceae) contains more than 67 species. It is quite widespread in the northern hemisphere, although *J. procera* Hochst. ex Endl. also grows southward along the rift mountains in East Africa in the southern hemisphere. The genus *Juniperus* is divided into three sections: *Caryocedrus* (one species, *J. drupacea* Labill.); *Juniperus* ("*Oxycedrus*, 9 or 10 species) and *Sabina* (the remaining, approximately 50 species) (Adams & Demeke, 1993; Adams, 1998, 2000, 2008). The flora of Algeria lists two sections and five *Juniperus* species; Sect. *Oxycedrus* (*J. communis* L., *J. oxycedrus* L.), and *Sabina* (*J. thurifera* L., *J. phonicea* L., *J. sabina* L.) (Quezel & Santa,

1963; Maire, 1967; Adams et al., 2003). All over the world plants from this genus have always been regarded as a well-known traditional remedy due to their numerous therapeutic properties, such as anti-inflammatory, diuretic, antiseptic (bacterial and fungal), hypoglycaemic, hypotensive, analgesic and abortifacient (Stassi et al., 1996; Milos & Radonic, 2000; Lesjak et al., 2011; Öztürk et al., 2011).

Thuriferous juniper (*Juniperus thurifera* L.) grows in the western part of the Mediterranean basin (France, Spain, Italia, Algeria and Morocco). It is a dioecious tree or shrub, with scale leaves and bluish black berries at maturity (Gauquelin, 2003). African subpopulations of *Juniperus thurifera* have been described as distinct species (*Juniperus africana* (Maire) Villar or more recently as a subspecies (*J. thurifera*

RESEARCH ARTICLE

subsp. *africana* (Maire) Romo and Boratynski (Romo & Boratynski, 2007). Genetic analysis also indicates that the Moroccan and Algerian subpopulations are genetically distinct and have been isolated from other subpopulations for several millennia (Terrab *et al.*, 2008).

Gender-related differences in growth and concentration of secondary metabolites have been documented in dioecious plants. Males usually grow faster than females, whilst females allocate more resources to reproduction and chemical defenses than males, hence their growth is reduced (McGowan *et al.*, 2004; Massei *et al.*, 2006; Cepeda-Cornejo & Dirzo, 2010). These physiological differences could also affect the secondary metabolism, as the production of volatile compounds (Gobbo-Neto & Lopes, 2007). In fact, a variation in the essential oil composition and phenolic component from male and female specimens was previously observed (Elmqvist *et al.* 1991; Riddle *et al.* 1996; Koztowska *et al.*, 2005; Massei *et al.*, 2006; Afsharypuor *et al.*, 2007; Lago *et al.*, 2008; Eiter *et al.*, 2010; Iszkuło *et al.*, 2011).

In recent years, essential oils and plant extracts have attracted a great deal of scientific interest due to their potential as a source of natural antioxidants and biologically active compounds, such as antibacterial, antifungal and insecticidal substances (Celiktaş *et al.*, 2007; Hammami *et al.*, 2011).

The hypothesis of this study was that the essential oil content and composition may be different in male and in female trees. Several works studied the chemical composition of the essential oils of *Juniperus thurifera* L. and their antimicrobials activity in different regions of the Mediterranean basin as France, Spain, the Pyrenees, the Corsicana and to Morocco (Hernandez *et al.*, 1987; Adams, 1999; Adams *et al.*, 2003; Barrero *et al.*, 2004; Achak *et al.*, 2008; Achak *et al.*, 2009; Mansouri *et al.*, 2010; Bahri *et al.*, 2013). But any work has not reported the effect of the sex of the trees on chemical composition of essential oils of *Juniperus thurifera* L.

Therefore, the objective of this study was to evaluate the variations in the yields, essential oil composition and their antibacterial activity within single male and female trees of Algerian thuriferous juniper (*Juniperus thurifera* L.).

Materials and Methods

Plant material

The leaves of male and female trees of *J. thurifera*, were collected from Aures Mountains (Algeria), at 1550 m of

altitude. A voucher specimen of each plant (sexes) is deposited in the herbarium of the Laboratory of Natural Resource Valorization, Faculty of Biology, Farhat Abbas University, Setif, Algeria.

The essential oils were extracted by hydrodistillation using Clevenger-type apparatus for three hours according to Tumen *et al.* (2010). The prepared volatile oils were dehydrated over anhydrous sodium sulphate and stored in sealed glass vials at 4-5°C prior to analysis.

Essential oil analysis

The essential oil analysis is performed on a chromatography's type Hewlett-Packard HP 7890 equipped with a capillary column DB-5 (length: 30 m, and 0.25 mm internal diameter, film thickness is 0.25 mm) coupled to a mass spectrometer (MS) type with a Hewlett Packard 5975 detector impact of electrons, 70 eV. The analytical conditions are as follows: injector temperature: 250°C, detector temperature: 280°C oven programming: 50 ° C for five min, 5°C / min from 50° to 300°C and isothermal 300°C for five min. The carrier gas is helium at a rate of one ml/min. Injector split mode 1:100. The GC analysis was carried out using an Agilent 6890N GC system equipped with a FID detector operated at a temperature of 300°C. To obtain the same elution order of peaks detected by GC/MS, simultaneous injection on the GC was performed using the same column and appropriate chromatographic conditions as those described for the GC/MS system.

Identification of the essential oil components was carried out by comparing their mass spectra and their KI (Kovats index) with those databases (Adams, 2001) and that established by the laboratory. The relative concentrations of the separated compounds based on percentage were computed from chromatograms obtained with the GC/FID system.

Antibacterial activity

Gram positive bacteria (*Staphylococcus aureus* ATCC 25923, *Staphylococcus aureus* clinical and methicillin-resistant *Staphylococcus aureus*) and Gram negative bacteria (*Pseudomonas aeruginosa* ATCC 27853, *Escherichia coli* ATCC 25922, *Escherichia coli* clinical, *Serratia marcescens*, *Enterobacter cloacae*, *Proteus permeri*, *Acinetobacter baumannii*, *Salmonella para*, *Salmonella spomb*, *Shigella* sp. and *Meningo* sp.) were used in this study.

The antimicrobial activity was tested by the agar-well diffusion method according to Mighri *et al.* (2010) with slight

RESEARCH ARTICLE

modifications. All bacterial cultures were first grown on Muller-Hinton at 37°C for 18-24 h prior to inoculation on to the nutrient agar. One or several colonies of similar morphology of the respective bacteria were transferred into API Suspension medium and adjusted to 0.5 McFarland turbidity standards with a densimat.

The inoculums of the respective bacteria were streaked onto Muller-Hinton agar plates using a sterile swab. A sterile filter disc (diameter 6 mm, Whatman paper No. 3) was placed. The disc was impregnated by the tested essential oils (30 µL/disc). The standard discs (6 mm diameter) of the antibiotics gentamycin (10 µg) were served as positive antibacterial control. The treated Petri dishes were placed at 4°C for 1–2 h and then incubated at 37°C for 18–24 h. Antimicrobial activity was evaluated by measuring the zone of growth inhibition around the discs after 24 h of incubation at 37°C. Each experiment was carried out in triplicate and the mean diameter of the inhibition zone was recorded.

The sensitivity to the different oils was classified by the diameter of the inhibition halos as follows: not sensitive (-) for diameter less than 8 mm; sensitive (+) for diameter 9-14 mm; very sensitive (++) for diameter 15-19 mm and extremely sensitive (+++) for diameter larger than 20 mm (Ponce *et al.*, 2003).

Statistical analysis

Variance analyses were used to test the significant difference among the results from the antibacterial assays (Minitab13). Means and standard errors (SE) of the samples were calculated. Three replicates were performed for each treatment.

Results and Discussion**Chemical composition of essential oils**

The hydrodistillation of leaf female trees of *J. thurifera* L. yielded 0.45% of essential oil (calculated on the basis of the mass of dry plant material), characterized by a strong odor and faint yellow color. However, the yield of the essential oil extracted from the leaves of male trees is 0.53% (v/w, dried material); characterized by a faint green color.

A total of 77 compounds were characterized and identified by gas chromatography (GC) and gas chromatography-mass spectrometry (GC/MS), representing more than 97.71% of the total oils (Table 1). The monoterpenes represented the main portion of the oils (≈80 % of the total essential oils), where monoterpene hydrocarbons

were most abundant, 63.2% in female trees and 58.53% in male trees. The main monoterpenes hydrocarbons were sabinene (32.11% in female trees, 31.16% in male trees) and α-pinene (13.04% in female trees, 7.84% in male trees), while the main oxygenated monoterpene was terpinen-4-ol (4.91% in female trees, 11.65% in male trees). Diterpene hydrocarbons were present in smaller amounts, 0.28% and 1.31%, oxygenated Diterpenes, 0.46% and 1.51%, and sesquiterpene hydrocarbons, 1.61% and 2.91% for essential oil of male and female trees respectively. Oxygenated sesquiterpenes were present in a high percentage in essential oil of male trees, amounting to 15.26% of the total essential oil content, while the essential oils of female trees contain 11.9% of the total essential oil content, with elemol as the main compound, 8.28% and 3.27% for essential oil of male and female trees respectively.

The concentrations of the oil constituents: α-thujene, α-pinene, α-phellandrene, p-cymene, linalyl acetate, Δ-amorphene, germacrene D-4-ol, and 4-epi-abietal were greater in the oil of the female tree than in the oil of the male tree. Conversely, the concentrations of α-terpinene, γ-terpinene, terpinene-4-ol, elemol, α-epi-cadinol and α-eudesmol were greater in the oil of the male tree than in the oil of the female tree. However, the concentration gradient trends for both female and male trees were similar for sabinene, myrcene, linalool, β-pinene, limonene, *cis*-sabinene hydrate terpinolene, α-terpineol,

Certain components detected in the female tree were not detected in the male trees, including β-phellandrene, isobutyl benzene, α-muurolole, epoxyde humulene II, valencene, α-muurolol, β-eudesmol and phytol. While the components, δ-2-carene, p-cymene-7-ol, verbanone, piperitone, cedrol, 1-epi-cubenol, cubenol and γ-gurjunene, were detected in the male tree and were not detected in the female trees.

Frequently, in dioecious plants, female plants allocate more resources to reproduction than male plants. Therefore it is expected that asymmetrical allocation to reproduction may lead to a reproduction-growth tradeoff, whereby female plants grow less than male plants, but invest more in defenses and thus experience lower herbivory than male plants (Cepeda-Cornejo & Dirzo, 2010). Typically, males allocate more resources to growth and females to high concentrations of secondary metabolites. Contrary to predictions, in this study the males had also higher yield in essential oils than females. This result is similar to that obtained by Asili *et al.* (2008a, 2010) for *Juniperus communis* subsp. *hemisphaerica*, *Juniperus oblonga* and *Juniperus foetidissima*, and

RESEARCH ARTICLE

Zheljazkov et al. (2013) for *Juniperus scopulorum* Sarg. The yield in essential oils obtained from the female leaves of *Juniperus sabina*, and *J. excelsa* subsp. *polycarpos* is higher to that obtained from male leaves (Asili et al., 2008b, 2010).

The results of the present study confirmed the prevalence of monoterpene hydrocarbons compared to other components. There are few previous reports on the phytochemical studies of the male and female essential oil of *Juniperus* species growing in other parts of the world indicated this results. Monoterpene hydrocarbons represented the most abundant constituents of the oil of both male and female leaves of shrubs of *J. communis* subsp. *hemisphaerica* (60.2 and 60.6% respectively, *J. oblonga*, (57.4 and 63.6%

respectively) (Asili et al., 2008a), *J. excelsa* subsp. *polycarpos* (81.57 and 66.42 % respectively) (Asili et al., 2008b; Emami et al., 2011), *J. chinensis* L. (60.2 and 60.6% respectively, *J. oblonga*, (37.2 and 42.7% respectively) (Afsharypuor et al., 2007), *J. sabina* (63.8 and 62.2% respectively) and *J. foetidissima* (75.9 and 68.3% respectively) (Asili et al., 2010).

A correlation between the chemical composition of the essential oil and the sex of the tree has been observed in several species of the genus *Juniperus* L. (Riddle et al., 1996; Afsharypuor et al., 2007; Zheljazkov et al., 2013) and are well known for other family (Lago et al., 2008; Eiter et al., 2010; Gourine et al., 2010; Iszkuło et al., 2011).

Table 1. Chemical composition of male and female *J. thurifera* essential oils.

Compound	Ki	Male (%)	Female (%)
α -thujene	924	1,8	2,06
α -pinene	932	7,86	13,04
fenchene	946	0,03	0,1
camphene	947	0,1	0,15
sabinene	973	31,16	32,11
β -pinene	976	1,18	1,56
myrcene	988	3,36	3,26
δ -2-carene	996	0,13	-
α -phellandrene	1006	0,06	1,06
α -terpinene	1015	2,72	1,47
p-cymene	1022	0,22	1,13
limonene	1027	1,7	1,1
β -phellandrene	1028	-	0,33
(Z)- β -ocimene	1035	0,05	0,11
(E)- β -ocimene	1045	0,21	0,4
γ -terpinene	1057	4,09	2,24
cis-sabinene hydrate	1069	1,97	1,51
terpinolene	1083	1,77	1,3
p-cymenene	1088	0,07	0,13
linalool	1098	3,14	3,77
cis-thujone	1104	0,17	0,24
trans-thujone	1114	0,17	0,55
cis-p-menth-2-en-1-ol	1123	0,85	0,38
α -comphlenal	1129	0,07	0,22
trans-p-menth-2-en-1-ol	1142	0,52	0,2
camphor	1144	0,05	0,14
E-tagetone	1147	0,1	0,18
terpinene-4-ol	1181	11,65	4,91
p-cymene-7-ol	1187	0,12	-
α -terpineol	1195	1,23	1
verbanone	1205	0,05	-
Trans-piperitol	1209	0,31	0,2
nerol 1 (80)	1224	0,08	0,12

RESEARCH ARTICLE

Compound	Ki	Male (%)	Female (%)
linalyl acetate	1250	1,32	3,16
piperitone	1255	0,39	-
pregeijerene	1280	0,59	0,14
bronyl acetate	1284	0,15	0,2
isobutyl benzene	1287	-	0,07
2,4-decadien-1-ol	1312	0,45	0,15
terpenyl acetate	1350	0,48	0,58
geranyle isobutyrate	1357	0,11	0,28
geranyle acetate	1377	0,21	0,63
β -caryophyllene	1418	0,04	0,1
α -humulene	1457	0,04	0,08
germacrene-D	1482	0,11	0,21
α -muurolene	1497	-	0,36
γ -cadinene	1515	0,07	0,12
Δ -amorphene	1517	0,1	1,6
elemol	1550	8,28	3,27
(E)-nerolidol	1560	0,3	0,33
germacrene D-4-ol	1579	0,29	1,24
Oxyde de Caryophyllene	1585	0,49	0,83
Aromadendr-9-ene	1592	0,13	0,12
cedrol	1596	0,16	-
humulene epoxyde II	1605	-	0,7
epi-cedrol	1613	0,2	0,73
valencene	1618	-	0,15
1-epi-cubenol	1627	0,36	-
γ -eudesmol	1630	0,26	0,98
α -epi-cadinol	1635	1,23	0,31
cubenol	1643	0,26	-
α -muurolol	1646	-	0,42
β -eudesmol	1649	-	0,05
α -eudesmol	1659	2,74	1,76
2,6-Dimethyl-1,5-heptadien-4-ol acetate	1666	0,17	0,58
gurjunene	1668	0,24	0,15
γ -gurjunene	1678	0,42	-
epi- α -bisabolol	1686	0,3	0,22
2-pentadecanone	1698	0,33	0,1
(Z,Z)-farnésol	1714	0,09	0,36
Cyclohexene, 1,5,5-trimethyl-6-(2-propenylidene)	1780	0,52	0,42
sandaracopimara-8(14), 15-diene	1962	0,13	0,59
manoyl oxide	1989	0,05	0,28
Abietatriène	2057	0,05	0,13
Abietatriène	2090	0,1	0,45
phytol	2109	-	0,14
4-epi-abietal	2299	0,41	1,27
Monoterpenes hydrocarbones		58,53	63,2
Oxygenated monoterpenes		21,57	16,84
Sesquiterpene hydrocarbons		1,61	2,91
Oxygenated sesquiterpenes		15,26	11,9
Diterpene hydrocarbons		0,28	1,31
Oxygenated Diterpenes		0,46	1,55

RESEARCH ARTICLE

Antibacterial activity

The antibacterial activity of both essential oils was evaluated by paper disc diffusion method against 14 bacteria. A zone of inhibition around the discs shows a bactericidal or bacteriostatic activity, while the absence of inhibition zone proved no effect of the tested oils against the microorganisms. We consider that essential oil has bacteriostatic action if the diameter of inhibition is >12 mm (Sağdaç, 2003). The results of the inhibition trials are reported in Table 2. The results showed that the oils inhibited the growth of bacterial strains produced a zone diameter of inhibition from 6 mm (no inhibition) to 17 mm. The maximum zone of inhibition was recorded against *Staphylococcus aureus* ATCC 25923 (17 mm) and *Escherichia coli* ATCC 25922 (16 mm). On the other hand, the oil was ineffective against *Pseudomonas aeruginosa* ATCC 27853.

The variance analyses showed significant difference among the results from the antibacterial assays (Table 3). The zone diameter of inhibition depended on susceptibility of the

tested bacteria, *Escherichia coli* ATCC 25922, *Escherichia coli* (clinical), *Staphylococcus aureus* ATCC 25923, *Staphylococcus aureus* (clinical), *Salmonella para*, *Salmonella spomb*, MRSA, are very sensitive again the essential oils, conversely, *Meningo* sp. *Serratia marcescens*, *Enterobacter cloacae* and *Proteus permeri* are not sensitive. *Shigella* sp. seems sensitive to the essential oils of *J. thurifera*. It is revealed that the *Escherichia coli* and *Staphylococcus aureus* was very sensitive to the essential oil of Moroccan *J. thurifera*, and two *Pseudomonas* strains proved resistant (Bahri et al., 2013).

The zone diameter of inhibition depended also on the essential oils type (from male or female trees). The essential oils of female trees showed a broad spectrum of antibacterial activity against Gram positive bacteria (*Staphylococcus aureus* ATCC 25923, *Staphylococcus aureus* clinical and MRSA) and Gram negative bacteria (*Escherichia coli* clinical, *Serratia marcescens*, *Proteus permeri*, *Meningo* sp. and *Shigella* sp).

Table 2. Inhibition diameter (mm) of leaf essential oil of *Juniperus thurifera*.

Bacteria	EO male leaves		EO female leaves		Gent.	
	means ± sd		means ± sd			
<i>Staphylococcus aureus</i> ATCC 25923	10.33±1.52	+	16±1	++	36	+++
<i>Staphylococcus aureus</i> (clinical)	12.66±1.15	+	13.33±1.15	+	35	+++
MRSA	10.33±0.57	+	13.66±1.52	+	35	+++
<i>Escherichia coli</i> ATCC 25922	15.66±0.57	++	7±0.57	-	27	+++
<i>Escherichia coli</i> (clinical)	0.00	-	11.33±0.57	+	23	+++
<i>Serratia marcescens</i>	0.00	-	10.33±0.57	+	0	-
<i>Enterobacter cloacae</i>	0.00	-	8.33±0.57	-	0	-
<i>Proteus permeri</i>	0.00	-	11.33±1,54	+	17	++
<i>Acinetobacter baumannii</i>	0.00	-	9.66±0.57	+	8	-
<i>Pseudomonas aeruginosa</i> ATCC 27853	0.00	-	0.00	-	32	+++
<i>Salmonella para</i>	12.66±0.57	+	9.66±0.57	+	20	+++
<i>Salmonella spomb</i>	13.66±0.57	+	11±0.57	+	16	++
<i>Meningo</i> sp.	0.00	-	10.66±0.57	+	0	-
<i>Shegnilla</i> sp.	8.33±0.57	-	10±1	+	27	+++

MRSA = Methicillin-resistant *Staphylococcus aureus*; Gent. = Gentamicine; EO = Essential oil; sd = standard deviation ;(-) = not sensitive; (+)= sensitive; (++) = very sensitive; (+++) = extremely sensitive.

Table 3. Variance analysis of two factors controlled.

	dl	SC	CM	F	P
Strains	13	476,488	36,653	58,09***	0,000
Essential oils	1	63,440	63,440	100,55***	0,000
Interaction	13	337,726	25,979	41,17***	0,000
Error	56	35,333	0,631		
Total	83	912,988			

RESEARCH ARTICLE

Conversely, the essential oils of male trees showed a broad spectrum of antibacterial activity against *Escherichia coli* ATCC 25922, *Staphylococcus aureus*, *Salmonella para*, and *Salmonella spomb*. This variability in the antibacterial activity of male and female *J. thurifera* essential oils is due to their different chemical composition.

The inhibitory action of the essential oil could be attributed to the occurrence of high proportions of monoterpenes and sesquiterpenes in the oil (Cakir et al., 2004). Antimicrobial properties of action might be related to these compounds which have a high potential in strongly inhibiting microorganism pathogens. The following components present are believed to play an important role as antibacterial agents, linalool, α -cadinol, globulol and viridiflorol, pulegone, epi- α -cadinol, terpinen-4-ol, germacrene A and *para*-methyl anisole corresponding to the amounts present in the essential oil (Hammami et al., 2011).

Conclusion

Analysis of the chemical composition of the essential oil of male and female *Juniperus thurifera* leaves has allowed identifying 77 compounds. The monoterpenes represented the main portion of the oils ($\approx 80\%$ of the total essential oils), The majority compounds are the sabinene, α -pinene, terpinene-4-ol, elemol, γ -terpinene and linalool. The essential oil was found to be active against all the bacterial strains except the *Pseudomonas aeruginosa* ATCC 27853.

The sex of the tree affected the chemical composition of essential oil and their antibacterial activity. Female trees grow less than male trees, but invest more in defenses than male trees. Consequently, the essential oil of female trees had the best bactericidal activity than the essential oils of female trees.

Acknowledgement

This study was supported in part by the Chemistry of heterocyclic compounds and carbohydrates Laboratory, Higher National School of Chemistry, Clermont Ferrand (France) and the Ministry of Higher Education and Scientific Research of the Algerian People's Democratic Republic.

References

- Achak N, Roman A, Alifriqi M, Adams PR. 2008. Effect of the leaf drying and sources on the essential oil composition of *Juniperus thurifera* L. Var. *africana* (Maire.) from Tensift Alhouz Marrekch (Morocco). J. Essent. Oil Res., 20: 200-204.
- Achak N, Roman A, Alifriqi M, Adams PR. 2009. Chemical studies of leaf essential oils of three species of *Juniperus* from Tensift Al Haouz-Marrakech Region (Morocco). J. Essent. Oil Res., 21(4): 337-341.
- Adams RP, Demeke T. 1993. Systematic relationships in *Juniperus* based on random amplified polymorphic DNAs (RAPDs). Taxon, 42: 553-572.
- Adams RR. 1998. The leaf essential oils and chemotaxonomy of *Juniperus* sect. *Juniperus*. Biochem. Syst. Ecol., 26: 637-645.
- Adams RP. 1999. Systematics of multi-seeded eastern hemisphere *Juniperus* based on leaf essentials and RAPD DNA fingerprinting. Biochem. Syst. Ecol., 27: 709-725.
- Adams RP. 2000. Systematics of *Juniperus* section *Juniperus* based on leaf essential oils and RAPD DNA fingerprinting. Biochem. Syst. Ecol., 28: 515-528.
- Adams RP. 2001. Identification of essential oils components by Gas Chromatography-Mass spectroscopy. - Illinois USA, Allured Publishing Corporation Carol Stream.
- Adams RP, Mumba LE, James SA, Pandey RN, Gauquelin T, Badri W. 2003. Geographic variation in the leaf oils and DNA Fingerprints (RAPDs) of *Juniperus thurifera* L. from Morocco and Europe. J. Essent. Oil Res., 15: 148-154.
- Adams RP. 2008. The Junipers of the World: The Genus *Juniperus*. - Victoria, BC, Canada: 2nd Ed. Trafford Publ.
- Afsharypour S, Rahiminezhad M, Ghaemmaghami L, Soleimani M, Khanmohammadi M, Afsharipour N. 2007. Essential oil constituents of leaves of the male and female shrubs of *Juniperus chinensis* L. from Isfahan. Iranian J. of Pharmaceutical Sci., 3(3): 177-180.
- Asili J, Emami SA, Rahimizadeh M, Fazly-Bazzaz BS, Hassanzadeh MK. 2008a. Chemical and antimicrobial studies of *Juniperus communis* subsp. *hemisphaerica* and *Juniperus oblonga* essential oils. J. Essent. Oil Bearing Plants, 11(1): 96-105.
- Asili J, Emami SA, Rahimizadeh M, Fazly-Bazzaz BS, Hassanzadeh MK. 2008b. Chemical and antimicrobial studies of *Juniperus excelsa* subsp. *excelsa* and *Juniperus excelsa* subsp. *polycarpus* essential oils. J. Essent. Oil Bearing Plants, 11(3): 292-302.
- Asili J, Emami SA, Rahimizadeh M, Fazly-Bazzaz BS, Hassanzadeh MK. 2010. Chemical and antimicrobial studies of *Juniperus sabina* L. and *Juniperus foetidissima* Willd. essential oils. J. Essent. Oil Bearing Plants, 13(1): 25-36.
- Bahri F, Harrak R, Achak N, Romane A. 2013. Chemical composition and antibacterial activities of the essential oils isolated from *Juniperus thurifera* L. var. *africana*. Nat. Prod. Res., 27(19): 1789-1794.
- Barrero AF, Quilez del Moral JF, Herrador MM, Akssira M, Bennamara A, Akkad S, Aitigri M. 2004. Oxygenated diterpenes and other constituents from Moroccan *Juniperus phoenicea* and *Juniperus thurifera* var. *africana*. Phytochemistry, 65: 2507-2515.

RESEARCH ARTICLE

- Cakir A, Kordali S, Zengin H, Izumi S, Hirata T. 2004. Composition and antifungal activity of essential oils isolated from *Hypericum hyssopifolium* and *Hypericum heterophyllum*. *Flav. Frag. J.*, 19(1): 62–68.
- Celiktas OY, Hames Kocabas EE, Bedir E, Verdar SO, Baser KHC. 2007. Antimicrobial activities of methanolic extract and essential oils of *Rosmarinus officinalis*, depending on location and seasonal variations. *Food Chemistry*, 100: 553-559.
- Cepeda-Cornejo V, Dirzo R. 2010. Sex-related differences in reproductive allocation, growth, defense and herbivory in three dioecious. *Neotropical Palms*, 5(3): e9824.
- Eiter LC, Fadamiro H, Setzer WN. 2010. Seasonal variation in the leaf essential oil composition of *Zanthoxylum clava-herculis* growing in Huntsville, Alabama. *Nat. Prod. Comm.*, 5(3): 457-460.
- Elmqvist T, Cates RG, Harper JK, Gardfjell H. 1991. Flowering in males and females of Utah willow, *Salix regida*, and effects on growth, tannins, phenolic glycosides and sugars. *Oikos*, 61: 65-72.
- Emami SA, Abedindo BF, Hassanzadeh-Khayyat M. 2011. Antioxidant activity of the essential oils of different parts of *Juniperus excelsa* M. Bieb. subsp. *excelsa* and *J. excelsa* M. Bieb. subsp. *polycarpus* (K. Koch) Takhtajan (Cupressaceae). *Iranian Journal of Pharmaceutical Research*, 10 (4): 799-810.
- Gauquelin T, Bertaudière V, Cambecedes C, Largier G. 2003. Le Jeneverier thurifère (*Juniperus thurifera* L.) dans les pyrenées: état de conservation et perspectives. *Acta. Bot. Barc.*, 49: 83-94.
- Gobbo-Neto L, Lopes NP. 2007. Plantas medicinais: Factores de influência no conteúdo de metabólitos secundários. *Quim. Nova*, 30: 374-381.
- Gourine N, Yousfi M, Bombarda I, Nadjemi B, Gaydou E. 2010. Seasonal variation of chemical composition and antioxidant activity of essential oil from *Pistacia atlantica* Desf. *Leaves. J. Am. Oil Chem. Soc.*, 87: 157–166.
- Hammami I, Triki MA, Rebai A. 2011. Chemical compositions, antibacterial and antioxidant activities of essential oil and various extracts of *Geranium sanguineum* L. flowers. *Arch. Appl. Sci. Res.*, 3 (3):135-144.
- Hernandez EG, Martinez CL, Villanova R. 1987. Determination by gas chromatography of terpenes in the berries of the species *Juniperus oxycedrus* L., *J. thurifera* L., and *J. sabina* L. *Journal of Chromatography*, 396: 416-420.
- Iszkulo G, Jasińska AK, Romo A, Tomaszewski D, Szymt J. 2011. The greater growth rate of male over female of the dioecious tree *Juniperus thurifera* only in worse habitat conditions. *Dendrobiology*, 66: 15-24.
- Kozłowska M, Bednorz L, Grzeškowiak M. 2005. Variability of *Juniperus communis* subsp. *communis* based on phenolics accumulation and glucosidase activity. *Dendrobiology*, 54: 29-33.
- Lago JHG, Romoff P, Fávero OA, Souza FO, Soares MG, Baraldi PT, Corrêa AG. 2008. Chemical composition of male and female *Baccharis trimera* (Less.) DC. (Asteraceae) essential oils. *Biochem. Syst. Ecol.* 36: 737-740.
- Lesjak MM, Beara IN, Orčić DZ, Anačkov GT, Balog KJ, Francišković MM, Neda M, Mimica-Dukić NM. 2011. *Juniperus sibirica* Burgsdorf. as a novel source of antioxidant and anti-inflammatory agents. *Food Chemistry*, 124: 850–856.
- Maire R, 1967. La flore de l'Afrique du Nord, Encyclopédie biologique. - Paris, France: Editions Paul Lechevalier.
- Mansouri N, Satrani B, Ghanmi M, El ghadraoui L, Aafi A, Farah A. 2010. Valorisation des huiles essentielles de *Juniperus thurifera* et de *Juniperus oxycedrus* du Maroc. *Phytothérapie*, 8: 166-170.
- Massei G, Watkins R, Hartley SE. 2006. Sex-related growth and secondary compounds in *Juniperus oxycedrus macrocarpa*. *Acta Oecologica*, 29: 135-140.
- McGowan GM, Joensalo J, Naylor R. 2004. Differential grazing of female and male plants of prostrate juniper (*Juniperus communis* L.). *Botanical Journal of Scotland*, 56(1): 39-54.
- Mighri H, Hajlaoui H, Akrouf A, Najjaa H, Neffati M. 2010. Antimicrobial and antioxidant activities of *Artemisia herba-alba* essential oil cultivated in Tunisian arid zone. *Comptes Rendus Chimie*, 13(3): 380–386.
- Milos M, Radonic A. 2000. Gas chromatography mass spectral analysis of free and glycosidically bound volatile compounds from *Juniperus oxycedrus* L. growing wild in Croatia. *Food Chemistry*, 68: 333-338.
- Öztürk M, Tümen I, Uğur A, Aydoğmus F, Topcu G. 2011. Evaluation of fruit extracts of six Turkish *Juniperus* species for their antioxidant, anticholinesterase and antimicrobial activities. *Journal of the Science of Food and Agriculture*, 91(5): 867–876.
- Ponce AG, Fritz R, del Valle C, Roura SI. 2003. Antimicrobial activity of essential oils on the native microflora of organic Swiss chard. *Lebensmittel- Wissenschaft und-Technologie*, 36: 679-684.
- Quezel P, Santa S. 1962. Nouvel Flore de l'Algérie et des régions désertiques méridionales. -Paris, France: Centre National de la Recherche Scientifique.
- Riddle RR, Charles A, Taylor JR, Kothmann MM, Huston JE. 1996. Volatile oil contents of ashe and redberry juniper and its relationship to preference by Angora and Spanish goats. *Journal of Range Manage*, 49:35-41.
- Romo A, Boratynski A. 2007. Nomenclatural note on *Juniperus thurifera* subsp *africana* (Cupressaceae). *Ann. Bot. Fennici*, 44: 72–75.
- Sağdaç O. 2003. Sensitivity of four pathogenic bacteria to Turkish thyme and oregano hydrosols. *LWT - Food Science and Technology*, 36(5): 467–473.
- Stassi V, Verekokidou E, Loukis A, Harvala C, Philianos S. 1996. The antimicrobial activity of the essential oils of four *Juniperus* species growing wild in Greece. *Flav. Fragr. J.*, 11: 71-74.
- Terrab A, Schönswetter P, Talavera S, Vela E, Stuessy TF. 2008. Range-wide phylogeography of *Juniperus thurifera* L., a presumptive keystone species of western Mediterranean vegetation during cold stages of the Pleistocene. *Mol. Phyl. Evol.*, 48: 94–102.
- Tumen I, Hafizoglu H, Pranovich A, Reunanen M. 2010. Chemical constituents of cones and leaves of cypress (*Cupressus sempervirens* L.) grown in Turkey. *Fresenius Environmental Bulletin*, 19 (10): 2268–2276.
- Zheljzakov VD, Astatkie T, Jeliazkova E. 2013. Year-round variations in essential oil content and composition of male and female juniper. *Hort Science*, 48(7): 883-886.